Surname	Centre Number	Candidate Number
Other Names		2



GCE A LEVEL - NEW

1400U50-1A



**BIOLOGY - A2 unit 5** 

**Practical Examination** 

Experimental Task TEST 1

TUESDAY, 4 APRIL 2017 2 hours

For Teacher's use only Award a mark of 0 or 1 for each of the following					
Correct measurement of volumes					
Syringes washed adequately					

For Examiner's use only				
Mark Awarded				
Total				

#### **ADDITIONAL MATERIALS**

In addition to this examination paper, you will require a calculator and a ruler.

#### **INSTRUCTIONS TO CANDIDATES**

Use black ink or black ball-point pen. Pencil may be used to draw tables and graphs. Write your name, centre number and candidate number in the spaces at the top of this page. Write your answers in the spaces provided in this booklet.

#### INFORMATION FOR CANDIDATES

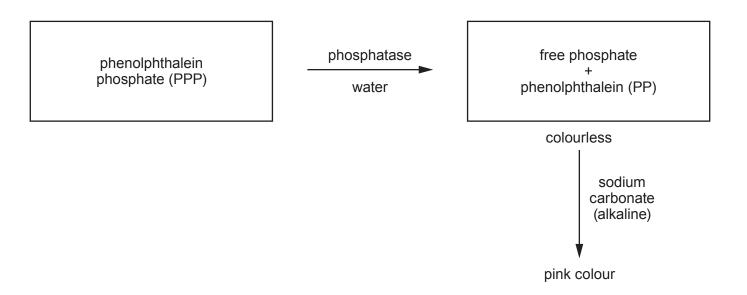
The total number of marks available for this task is 20.

Your teacher will directly assess your practical skills.

The number of marks is given in brackets at the end of each question or part question.

You are reminded of the necessity for orderly presentation in your answers.

This practical involves the action of the enzyme **phosphatase** which acts on substrates to release phosphate groups. In this investigation the substrate for the enzyme is phenolphthalein phosphate (PPP). Phosphatase reacts with PPP to produce phenolphthalein (PP) and a free phosphate group as shown below.



Both products are colourless in neutral and acid conditions.

When alkaline sodium carbonate is added, the phenolphthalein turns pink and the phosphatase is inactivated.

You are provided with an extract from mung beans which contains the enzyme phosphatase.

In this investigation you are going to determine how the production of phenolphthalein changes during the course of the reaction.

The experiment will be carried out at room temperature and pH7.

### You are given:

- 1. 2 syringes each with a volume of 5 cm<sup>3</sup>
- 2. 2 syringes each with a volume of 1 cm<sup>3</sup>
- 3. Enzyme extract (approximately 20 cm<sup>3</sup>) in pH7 buffer
- 4. 15 test tubes
- 5. Solution of sodium carbonate (approximately 20 cm<sup>3</sup>)
- 6. Solution (approximately 20 cm<sup>3</sup>) of phenolphthalein phosphate (the substrate)
- 7. Stop clock
- 8. Colour chart of phenolphthalein solutions (page 3)
- 9. Glass rod
- 10. 3 beakers each with a volume of 100 cm<sup>3</sup>
- 11. 250 cm<sup>3</sup> beaker of distilled water
- 12. 250 cm<sup>3</sup> beaker for waste
- 13. Test tube rack
- 14. Paper towels
- 15. Safety goggles

Turn over.

## YOUR TEACHER WILL BE OBSERVING YOUR EXPERIMENTAL TECHNIQUE.

[2]

#### Method

- **1.** Use one syringe to place 1 cm<sup>3</sup> of the sodium carbonate solution into each of 5 test tubes. Do not use this syringe again.
- 2. Use a clean syringe to add 5 cm<sup>3</sup> of substrate (PPP) to a 100 cm<sup>3</sup> beaker.
- **3.** Use a **different** syringe to add 5 cm<sup>3</sup> of enzyme extract to the substrate. Mix and **immediately** start the stop clock.
- **4. After 1 minute** and then at 1 minute intervals, use a 1 cm<sup>3</sup> syringe to remove 1 cm<sup>3</sup> of the enzyme and substrate mixture, and add it to one of the test tubes which contains sodium carbonate solution.
- **5.** Before taking the next sample, fill the syringe with distilled water and discard the waste into the beaker provided.
- **6.** Use the colour chart provided below to estimate the concentration of phenolphthalein (PP) in each of your 5 test tubes, in mol dm<sup>-3</sup>. Record your results clearly.
- 7. Repeat the whole investigation twice to give three readings for each time interval.

			Colo	ur ch	art foi	r phen	olpht	halein	solut	tions		
Ľ.												
atic												
Soncentration of PP	0-6	0-6		0-6		0-6		0-6		10-5	10-5	10-5
PP	x 1	х д		x 7 dq		x 7		x dm		0 × 0	x mb	, × 0
G C	0.0 mol	1.0 mol		2.0 mol		3.0 mo		5.0 mol		1.20 mol	2.50 mol (	7.50 mol

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(a) Below, record your results accurately and clearly.

Examiner only

[3]

[9]

(b) Plot the results from your table on the grid below.

(c)	(i)	With reference to your graph discuss the consistency of your readings. [2]
	(ii)	Suggest <b>two</b> sources of inaccuracy in this investigation and an improvement for each. [2]
		Inaccuracy 1:
	•••••	Improvement
		Inaccuracy 2:
		Improvement
(d)	Sugg	gest how the experiment could be modified to investigate the effect of pH on the rity of phosphatase. [2]

20

## **END OF PAPER**

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